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**SYNTHESIS OF NEW NUCLEOSIDE ANALOGUES COMPRISING A
GEMINAL DIFLUOROCYCLOPROPANE MOIETY AS POTENTIAL
ANTIVIRAL/ANTITUMOR AGENTS**

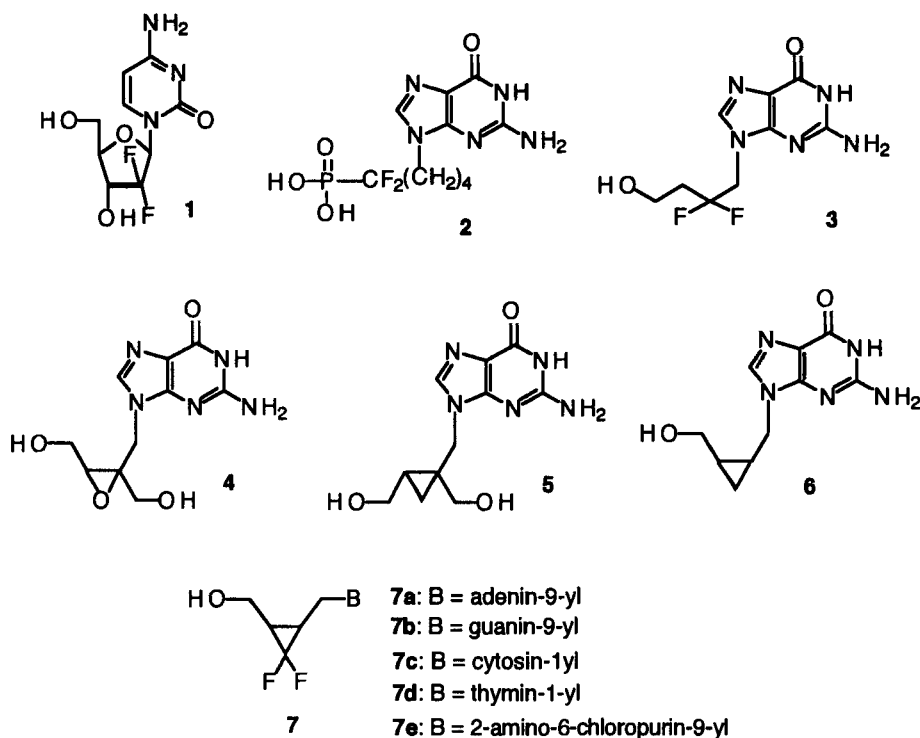
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ABSTRACT. Geminal difluorocyclopropane analogues of nucleosides **7a** - **7e** were synthesized. Compounds **7a** and **7c** - **7e** were obtained by alkylation of nucleic acid bases or their appropriate precursors with (*cis*)-1-benzyloxymethyl-2-bromomethyl-3,3-difluorocyclopropane (**8**). Analogue **7b** was prepared by hydrolysis of 2-amino-6-chloropurine derivative **7e**. Compounds **7a** - **7d** did not exhibit any antiviral activity against HCMV, HSV-1, HSV-2, EBV, VZV, HBV and HIV-1 or antitumor effects against murine leukemia L1210, mouse tumors PO3 or C38 and human tumor H15.

Nucleoside analogues carrying a fluorine substitution in the heterocyclic or carbohydrate moiety provided a plethora of biologically active compounds¹⁻³. Nevertheless, examples of biologically active analogues of nucleosides or nucleotides comprising a difluoromethylene function CF₂ are rather scant. The most notable example is 2'-deoxy-2',2'-difluorocytidine (**1**, gemcitabine, Gemzar), a powerful antitumor agent.⁴ It was also reported that geminal difluoromethylene moiety is a close steric and electronic mimic of oxygen atom.¹ Indeed, 9-(5',5'-difluoro-5'-phosphonopentyl)guanine **2**, an acyclic analogue of dGMP, is an effective inhibitor of purine nucleoside phosphorylase⁵. This concept was also applied in the area of acyclic nucleoside analogues. Thus, difluoromethylene mimic of antiherpetic drug acyclovir (Zovirax), compound **3**, was found⁶ to have antiherpetic activity albeit lower than the parent drug.

Further related to this work, the oxirane analogue⁷ **4** also exhibited some anti-herpesvirus potency. However, this compound was not completely stable under the conditions of antiviral assays. More stable cyclopropane analogues^{7,8} **5** and **6** related to oxirane **4** exhibited potent effects against herpesviruses α .

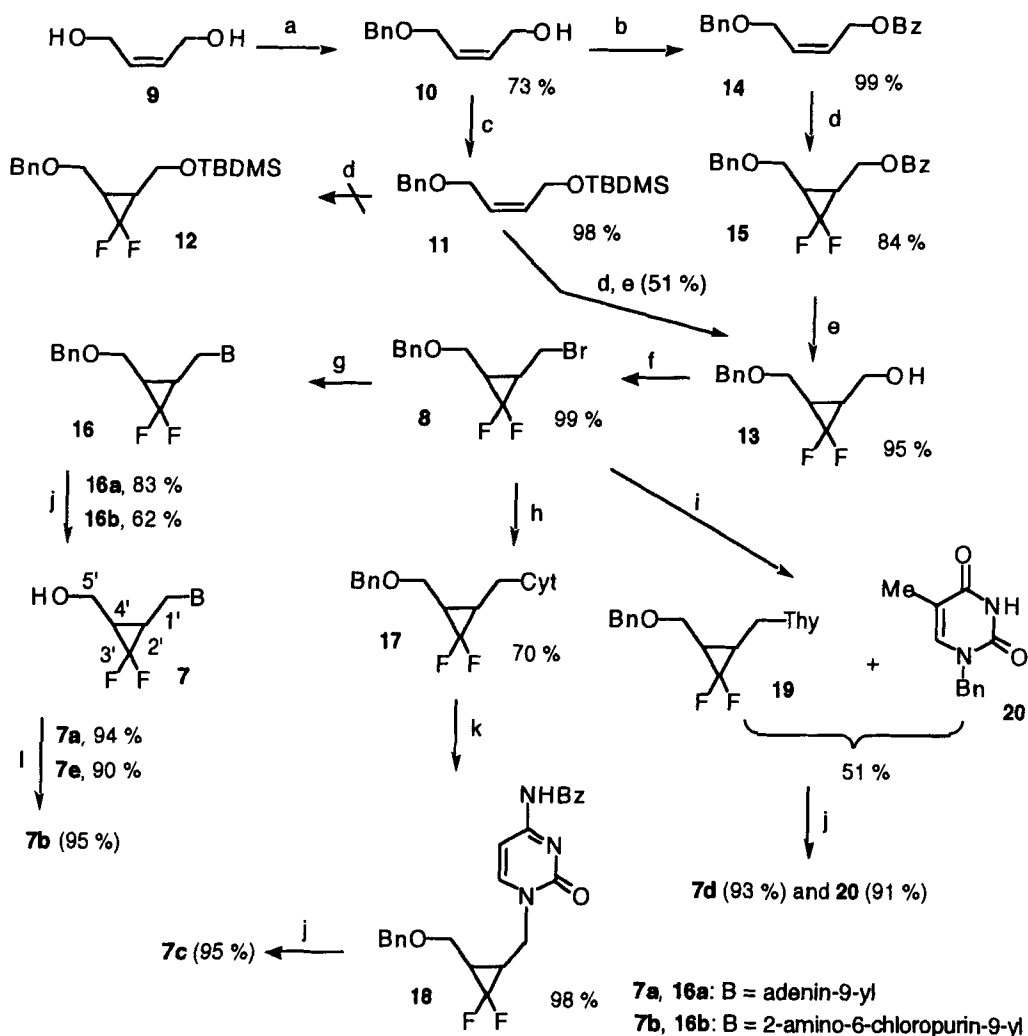


All these findings formed the basis for an assumption that cyclopropane analogues of acyclovir with a geminal difluoromethylene moiety such as **7a** - **7d** will be more stable than the corresponding oxirane derivatives such as **4**. In addition, geminal difluorocyclopropane moiety will be a close steric and electronic mimic of oxirane ring. In this communication we describe the synthesis and investigation of antiviral/antitumor activity of geminal difluorocyclopropane analogues **7a** - **7d**.

RESULTS AND DISCUSSION

At the outset, elaboration of an alkylating reagent which could be employed for alkylation of any desired nucleic acid base or suitable precursor was considered as the most convenient approach for synthesis of analogues **7a** - **7d**. Such an agent, difluorocyclopropane derivative **8**, was obtained as follows (Scheme 1). Commercially available (*cis*)-2-butene-1,4-diol (**9**) was converted to a monobenzyl ether **10** by reaction with sodium hydride followed by treatment with benzyl bromide and tetrabutylammonium iodide (NBu₄I) in tetrahydrofuran⁹ (51 % yield). Using a 200 % molar excess of **9** the yield of **10** was increased to 73 %. These results compare favorably with the previously described two-step method^{10,11} starting from cyclic benzaldehyde acetal of **9**. Compound

Scheme 1

a. 1. NaH, THF. 2. Bu₄NI, BnBr.

b. BzCl, pyridine.

c. TBDMS, imidazole, DMF.

d. ClF₂CCO₂Na, diglyme, 180°C.e. K₂CO₃, MeOH - H₂O.f. CBr₄, Ph₃P, CH₂Cl₂.g. B-H, K₂CO₃, DMF, Δ.h. 1. N⁴-Acetylcytosine, K₂CO₃, DMF, Δ. 2. H₂O, Δ.

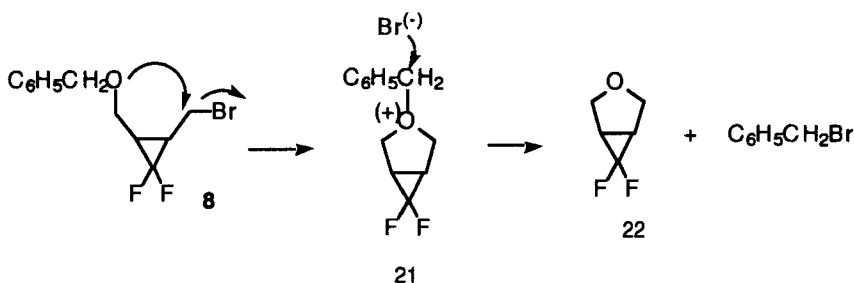
i. 2,4-Bis-O-TMS-5-methylpyrimidine, MeCN, Δ, 7 days.

j. 1. BCl₃, CH₂Cl₂, -78°C. 2. NH₃, MeOH or NaHCO₃, MeOH.k. Bz₂O, EtOH, Δ.l. 1. 80 % HCO₂H, Δ. 2. NH₃, MeOH.

10 was converted to a fully protected diol **11** by reaction with tert.-butylsilyldimethylsilyl chloride (TBDMSCl) and imidazole¹² in DMF. Attempted addition of difluorocarbene generated from sodium chlorodifluoroacetate in diglyme at 180°C according to the described procedure¹³ (the reported yield was 95 %) failed to give any difluorocyclopropane intermediate **12**. The crude reaction product contained only benzyl derivative **13** contaminated with 5 - 10 % of the chlorodifluoroacetate ester (tentative structure). Treatment of the reaction mixture with K₂CO₃ in aqueous methanol gave a 51 % yield of **13**. This result showed that the TBDMS group did not survive the conditions of the difluorocarbene addition. Therefore, compound **10** was converted to benzoate **14**. Addition of difluorocarbene afforded smoothly intermediate **15** in 84 % yield. Debenzylation with K₂CO₃ in aqueous methanol furnished compound **13** (95 %). The latter was transformed to the corresponding bromo derivative **8** by reaction¹⁴ with CBr₄ and P(C₆H₅)₃ in CH₂Cl₂ (99 %).

Alkylation of nucleic acid bases with reagent **8** was performed using procedures previously elaborated for synthesis of methylenecyclopropane analogues.^{15,16} Thus, reaction of adenine with **8** using K₂CO₃ in DMF at 85°C gave intermediate **16a** (83 %). Debenzylation was effected with BCl₃ in CH₂Cl₂ and subsequent treatment with NH₃ in methanol as described for an allenic analogue¹⁷ to afford compound **7a** (94 %). In a similar vein, alkylation of 2-amino-6-chloropurine with reagent **8** led to protected derivative **16b** (62 %). After debenzylation, analogue **7e** was obtained in 90 % yield. Hydrolysis of **7e** with 80 % formic acid followed by treatment with NH₃ in MeOH¹⁵ gave guanine analogue **7b** (95 %). Alkylation with reagent **8** was also employed in the pyrimidine series. Thus, reaction of N⁴-acetylcytosine with **8** under the conditions described for adenine analogue **16a** gave, after deacetylation, protected intermediate **17** in 70 % yield. Attempted debenzylation with BCl₃ was not successful because of a poor solubility of **17** in CH₂Cl₂. Therefore, compound **17** was benzoylated using benzoic anhydride in refluxing ethanol as described for N⁴-benzoylcytallene¹⁸ to give N⁴-benzoyl derivative **18** (98 %). The latter was smoothly debenzylated using BCl₃ and subsequent debenzylation furnished the desired analogue **7c** in 95 % yield. Alkylation of thymine with **8** also followed the procedure described for synthesis of the corresponding methylenecyclopropane analogues.¹⁶ The 2,4-bis-O-trimethylsilyl-5-methylpyrimidine was refluxed with reagent **8** in acetonitrile for 7 days to give a mixture of protected analogue **19** and N¹-benzylthymine (**20**) in the ratio of 1.5 : 1 and 51 % yield, which were inseparable by column chromatography on silica gel. Deprotection with BCl₃ and subsequent chromatography afforded N¹-benzylthymine (**20**) in 91 % yield (the N-benzyl group was apparently stable) and analogue **7d** (93 %). It should be noted that a similar N-benzylation was observed during alkylation of thymine with 4'-benzyloxy-3'-

Scheme 2



benzyloxymethylbutyl bromide under basic catalysis¹⁹. In our case, this side-reaction is best rationalized in terms of Scheme 2. The cyclic oxonium intermediate **21** formed from **8** is attacked by bromide ion to give benzyl bromide and bicyclic derivative **22**. Benzyl bromide then competes with reagent **8** in alkylation of the silylated thymine. Alternately, intermediate **21** may also serve as a benzylating agent.

The structures of analogues **7a** - **7e** were fully supported by spectroscopic data. The UV spectra are in agreement with the assignment as N^9 -isomers for purines **7a**, **7b** and **7e** or N^1 -isomers for pyrimidines **7c** and **7d**. No appreciable amounts of other isomers were detected in appropriate crude products.

Analogues **7a** - **7d** were tested against the following viruses: HCMV, HSV-1, HSV-2, EBV, VZV, HBV and HIV-1. The details of the assays were described previously.¹⁵ The antitumor zone assays²⁰ were performed with murine leukemia L1210, mouse tumors PO3 or C38 and human tumor H15. Neither of the analogues exhibited a significant antiviral effect, antitumor activity or cytotoxicity in any of these assays at the highest concentration tested ($100\text{ }\mu\text{M}$, EBV/H-1 $50\text{ }\mu\text{M}$ and HBV $10\text{ }\mu\text{M}$). The lack of biological potency can be explained by a poor substrate ability toward relevant viral or cellular kinases or that the phosphorylated metabolites are poor inhibitors of viral or cellular polymerases. Adenine derivative **7a** was not a substrate for adenosine deaminase from calf intestine.

EXPERIMENTAL SECTION

General Methods. See¹⁵. The NMR spectra were determined at 300 or 400 MHz (^1H), 75 or 100 MHz (^{13}C) and 282 or 376 MHz (^{19}F) in CD_3SOCD_3 unless stated otherwise. For fast atom bombardment mass spectra (FAB-MS) thioglycerol (TG) matrix was used. (*cis*)-4-Benzoyloxy-2-buten-1-ol (**10**). Sodium hydride (50 % in mineral oil, 5.00 g, 0.11 mol) was added in five equal portions every 5 minutes with stirring and ice-cooling

under N₂ to a solution of (*cis*)-2-buten-1,4-diol (**9**, 9.25 g, 0.11 mol) in THF (100 mL). The suspension was then stirred for 3 h at room temperature. The (Bu)₄Ni (387 mg, 1.1 mmol) followed by benzyl bromide (12.48 mL, 0.11 mol) were then added and the mixture was stirred for another 16 h. The insoluble portion was filtered off using a Celite pad and it was washed with CH₂Cl₂ (3 x 20 mL). The organic phase was evaporated and the residue was chromatographed on a silica gel column using hexane - ethyl acetate (7 : 3 → 2 : 3) to give product **10** as an oil (9.36 g, 50 %).

In another experiment (0.3 mol scale based on benzyl bromide), using 0.9 mol of diol **9** product **10** was obtained in 73 % yield after distillation (bp 96-98°C/0.2 torr). ¹H NMR (CDCl₃) δ 2.62 (br s, 1 H, OH), 4.13 (dd, 2 H, ³J = 6.0 Hz, ⁴J = 0.6 Hz) and 4.07 (dd, 2 H, ³J = 6.0 Hz, ⁴J = 0.9 Hz, =CHCH₂O), 4.52 (s, 2 H, CH₂C₆H₅), 5.67-5.84 (m, 2 H, ³J = 6.0 Hz, ⁴J = 1.2 Hz, HC=CH), 7.26-7.39 (m, 5 H, C₆H₅).

(cis)-1-Benzoyloxy-4-(tert-butyldimethyl)silyloxy-2-butene (11). DMF was evaporated in vacuo from a solution of (*cis*)-4-benzyloxy-2-buten-1-ol (**10**, 9.36 g, 52.5 mmol) and imidazole (5.36 g, 78.8 mmol) in DMF (20 mL). The residue was redissolved in DMF (30 mL) and tert-butyldimethylsilyl chloride (9.50 g, 63 mmol) was added with stirring at room temperature. The mixture was stirred for 16 h. Volatile components were removed in vacuo and the residue was partitioned between hexane (150 mL) and water (50 mL). The aqueous phase was extracted with hexane (20 mL). The combined hexane phases were washed with HCl (1 M, 50 mL), water (50 mL), saturated NaHCO₃ (50 mL), water (50 mL), brine (50 mL) and they were dried (Na₂SO₄). After evaporation, the residue was chromatographed on a silica gel column using hexane - ethyl acetate (98 : 2 → 95 : 5) to afford silyl ether **11** (15.05 g, 98 %) as an oil. ¹H NMR (CDCl₃) δ 0.08 (s, 6 H, Me₂^tBuSi), 0.92 (s, 9 H, ^tBu), 4.00 (d, 2 H, ³J = 6.0 Hz, CH₂OSi), 4.24 (d, 2 H, ³J = 5.7 Hz, CH₂OBn), 4.53 (s, 2 H, CH₂Ph), 5.62-5.79 (m, 2 H, apparent ³J = 5.9 Hz, ⁴J = 0.9 Hz, H alkene), 7.26-7.39 (m, 5 H, Ph). ¹³C NMR (CDCl₃, 75 MHz): -5.21 (Me₂^tBuSi), 18.31 (SiCMe₃), 25.91 (CMe₃), 59.56 (CH₂OSi), 65.83 (CH₂OBn), 126.81 and 127.63 (C alkene); benzyl: 72.22 (CH₂), 127.80 (C_{ortho}), 128.40 (C_{meta}), 132.90 (C_{para}), 138.21 (C_{ipso}).

(cis)-1-Benzoyloxy-4-benzyloxy-2-butene (14). Benzoyl chloride (2.90 mL, 25 mmol) was added into a solution of (*cis*)-4-benzyloxy-2-buten-1-ol (**10**, 1.78 g, 10 mmol) in pyridine (10 mL) at room temperature with stirring which was continued for 16 h. Water (3 mL) was added and the resulting mixture was stirred for 24 h. Volatile components were removed in vacuo and the residue was partitioned between hexane (150 mL) and water (50 mL). The organic phase was washed with HCl (1 M, 2 x 50 mL), water (50 mL), Na₂CO₃ (5 %, 2 x 50 mL), brine (50 mL) and it was dried (Na₂SO₄). After evaporation, the residue was chromatographed on a silica gel column using hexane - ethyl acetate (9 : 1) to give product **14** as an oil (2.80 g, 99 %). IR (neat) 1715 (C=O) cm⁻¹. ¹H NMR (CDCl₃)

δ 4.23 (d, 2 H, $^3J = 4.8$ Hz, CH_2OBn), 4.57 (s, 2 H, CH_2Ph), 4.91 (d, 2 H, $^3J = 5.7$ Hz, CH_2OBz), 5.83-5.97 (m, 2 H, alkene H), 7.27-7.42 (m, 5 H, OCH_2Ph), 7.42-7.50 (m, 2 H) and 7.55-7.62 (m, 1 H) and 8.06-8.11 (m, 2 H, Bz). ^{13}C NMR 60.80 (CH_2OBz), 65.75 (CH_2OBn), 126.73 and 127.78 (C alkene); benzyl: 72.48 (CH_2), 127.85 (Cortho), 128.41 (Cmeta), 131.08 (Cpara), 138.03 (Cipso); benzoyl: 128.47 (Cortho), 129.65 (Cmeta), 130.12 (Cpara), 133.04 (Cipso), 166.39 (C=O).

(*cis*)-1-Benzoyloxymethyl-3-benzoyloxymethyl-2,2-difluorocyclopropane

(15). A suspension of sodium chlorodifluoroacetate (7.62 g, 50 mmol) in diglyme (17 mL) was added into a solution of (*cis*)-1-benzoyloxy-4-benzoyloxy-2-butene (**14**, 1.41 g, 5 mmol) in diglyme (5 mL) heated at 180°C (bath temperature) with stirring over 4.5 h using a syringe pump. The mixture was then refluxed for 3 h. After cooling, the solvent was evaporated in vacuo. Hexane (150 mL) was added to the residue, the insoluble portion was filtered off using a Celite pad and it was washed with the same solvent (3 x 10 mL). The hexane phase was washed with water (50 mL) and brine (50 mL) whereupon it was concentrated in vacuo. Water (20 mL) was added followed by solid KMnO_4 in portions (total 510 mg) at 0°C with stirring until the pink color persisted for 1.5 h. The solids were filtered off using a Celite pad and they were washed by sonication with hexane (8 x 25 mL). The organic phase was dried (Na_2SO_4), evaporated and the residue was chromatographed on silica gel using hexane - ethyl acetate (95 : 5 \rightarrow 9 : 1) to give difluorocyclopropane derivative **15** (1.39 g, 84 %) as an oil. IR (neat) 1740 (C=O), 1495, 1295, 1115, 1090, 1040, 730 cm^{-1} ; ^1H NMR (CDCl_3) δ 2.09-2.32 (m, 2 H, apparent $J = 8.0$ Hz, $J = 1.6$ Hz, cyclopropyl H), 3.69-3.80 (m, 2 H, apparent $J = 8.0$ Hz, $J = 1.6$ Hz, CH_2OBn), 4.44-4.60 (m, 2 H, apparent $J = 2.4$ Hz, CH_2OBz), 4.56 (AB, 2 H, $J_{\text{AB}} = 12.0$ Hz, OCH_2Ph), 7.30-7.40 (m, 5 H, OCH_2Ph), 7.44-7.50 (m, 2 H) and 7.58-7.63 (m, 1 H) and 8.05-8.09 (m, 2 H, Bz). ^{13}C NMR 24.99 (t, $^2J_{\text{F}} = 10.4$ Hz) and 26.44 (t, $^2J_{\text{F}} = 10.4$ Hz, cyclopropyl CH), 59.55 (d, $^3J_{\text{F}} = 6.0$ Hz, CH_2OBz), 64.00 (d, $^3J_{\text{F}} = 5.2$ Hz, CH_2OBn), 114.25 (dd, $^1J_{\text{F}} = 288.1$ Hz, $^1J_{\text{F}} = 281.4$ Hz, CF_2); benzyl: 74.12 (CH_2), 128.88 (Cortho), 128.97 (Cpara), 129.55 (Cmeta), 138.68 (Cipso); benzoyl: 129.61 (Cortho), 130.81 (Cmeta), 130.87 (Cpara), 134.87 (Cipso), 167.45 (C=O). ^{19}F NMR -124.60 (dt, $^2J_{\text{F}} = 163.4$ Hz, $^3J_{\text{cis-H}} = 13.0$ Hz), -150.19 (d, $^2J_{\text{F}} = 164.9$ Hz). EI-MS 332 (M, 0.9), 227 (M - Bz, 3.3), 225 (M - OBn, 6.3), 210 (M - OBz - H, 7.9), 206 (M - OBn - F, 17.3), 122 (BzOH, 11.9), 105 (Bz, 100.0), 91 (Bn, 89.5), 77 (Ph, 27.9); CI-MS 333 (M + H, 7.7), 225 (M - OBn, 4.7), 211 (M - OBz, 15.7), 105 (Bz, 25.9), 91 (Bn, 100); HRMS: Calcd. for $\text{C}_{19}\text{H}_{18}\text{F}_2\text{O}_3$ (M): 332.1224. Found: 332.1221. Anal. Calcd. for $\text{C}_{19}\text{H}_{18}\text{F}_2\text{O}_3$: C, 68.67; H, 5.46; F, 11.43. Found: C, 68.84; H, 5.60; F, 11.51.

(*cis*)-1-Benzoyloxymethyl-2,2-difluoro-3-hydroxymethylcyclopropane (13).

Method A. From Benzoate 15. A mixture of compound **15** (15.87 g, 47.8 mmol)

and K_2CO_3 (6.60 g, 47.8 mmol) in $\text{MeOH} - \text{H}_2\text{O}$ (9 : 1, 160 mL) was stirred at room temperature for 16 h. It was then partitioned between ethyl acetate and water (200 mL each). The aqueous phase was extracted with ethyl acetate (3 x 100 mL). The combined organic phase was washed with Na_2CO_3 (5 %, 3 x 100 mL), water (2 x 100 mL), brine (70 mL) and it was dried (Na_2SO_4). After evaporation, the residue was chromatographed on silica gel using hexane - ethyl acetate (85 : 15 \rightarrow 7 : 3) to give product **13** (10.36 g, 95 %) as an oil. The ^1H and ^{13}C NMR spectra corresponded to those described¹³.

Method B. From TBDMS Derivative 11. The reaction was performed as described¹³ using compound **11** (13.57 g, 46.39 mmol) and sodium chlorodifluoroacetate (70.74 g, 464 mmol) to give product **13** contaminated by its chlorodifluoroacetate (tentatively assigned, 5-10 %) after chromatography on silica gel using hexane - ethyl acetate as eluent as described above. Treatment of the crude product (7.40 g) with K_2CO_3 in $\text{MeOH} - \text{H}_2\text{O}$ as described above afforded compound **13** (5.35 g, 51 %). The ^1H NMR spectrum was identical with that of the product **13** prepared by method A.

(cis)-1-Benzyloxymethyl-2-bromomethyl-3,3-difluorocyclopropane (8).

Triphenylphosphine (4.31 g, 16.4 mmol) was added to a solution of compound **13** (2.53 g, 11.1 mmol) and CBr_4 (6.19 g, 18.65 mmol) in CH_2Cl_2 (26 mL) at room temperature with stirring in five equal portions and 5 min intervals. After evaporation, the residue was passed through a short column of silica gel (2.5 x 10 cm) eluted with CH_2Cl_2 (200 mL), to give a crude bromo derivative **8**. This product was rechromatographed using hexane - ethyl acetate (98 : 2 \rightarrow 9 : 1) to give compound **8** (3.18 g, 98.5 %) as an oil. IR (neat) 1480, 1300, 1220, 1100, 1040, 750, 710 cm^{-1} ; ^1H NMR (CDCl_3) δ 2.13 (ddtd, 1 H, $^3J_{\text{cis-F}} = 13.6$ Hz, $^3J_{\text{cis-H}} = 11.0$ Hz, $^3J_{\text{H}} = 7.8$ Hz, $^3J_{\text{trans-F}} = 1.9$ Hz, CHCH_2Br), 2.25 (td, 1 H, $^3J_{\text{cis-F}} = 3J_{\text{cis-H}} = 12.0$ Hz, $^3J_{\text{H}} = 8.4$ Hz, $^3J_{\text{H}} = 7.4$ Hz, $^3J_{\text{trans-F}} = 1.5$ Hz, CHCH_2OBn), 3.45 (tm, 1 H, $^2J = ^3J = 9.8$ Hz) and 3.53 (ddd, 1 H, $^2J_{\text{H}} = 10.8$ Hz, $^3J_{\text{H}} = 7.6$ Hz, $^4J_{\text{F}} = 3.2$ Hz, CH_2OBn), 3.70 (AB x q, 2 H, $J_{\text{AB}} = 12.4$ Hz, $^3J_{\text{H}} = ^4J_{\text{F}} = 1.6$ Hz, CH_2Br), 4.56 (AB, 2 H, $J_{\text{AB}} = 12.0$ Hz, OCH_2Ph), 7.32-7.43 (m, 5 H, Ph). ^{13}C NMR 25.36 (d, $^3J_{\text{F}} = 5.9$ Hz, CH_2Br), 28.15 (t, $^2J_{\text{F}} = 10.0$ Hz) and 28.62 (t, $^2J_{\text{F}} = 10.8$ Hz, cyclopropyl CH), 63.35 (d, $^3J_{\text{F}} = 5.2$ Hz, CH_2OBn), 114.54 (dd, $^1J_{\text{F}} = 291.1$ Hz, $^1J_{\text{F}} = 282.2$ Hz, CF_2); benzyl: 74.16 (CH_2), 128.91 (C_{ortho}), 129.09 (C_{para}), 129.66 (C_{meta}), 138.62 (C_{ipso}). ^{19}F NMR -124.36 (dt, $^2J_{\text{F}} = 163.4$ Hz, $^3J_{\text{cis-H}} = 12.2$ Hz), -151.89 (d, $^2J_{\text{F}} = 163.4$ Hz). EI-MS 291 and 289 ($\text{M} + \text{H}$, 0.6, 0.5), 211 ($\text{M} - \text{Br}$, 1.6), 191 ($\text{M} - \text{Br} - \text{F} - \text{H}$, 0.3), 181 (3.3), 172 and 170 ($\text{M} - \text{CH}_2\text{OBn} + \text{H}$, 0.5, 0.5), 161 (1.5), 105 (6.1), 91 (Bn, 100); CI-MS 293 and 291 ($\text{M} + \text{H}$, 6.0, 7.4), 211 ($\text{M} - \text{Br}$, 6.2), 181 (9.7), 91 (Bn, 100). HRMS Calcd. for $\text{C}_{12}\text{H}_{13}^{79}\text{BrF}_2\text{O}$ (M) 290.0118. Found: 290.0113.

(cis)-9-[(2-Benzyloxymethyl-3,3-difluorocyclopropyl)methyl]adenine

(16a). A mixture of adenine (405 mg, 3 mmol), compound **8** (582 mg, 2 mmol) and

K₂CO₃ (1.1 g, 8 mmol) in DMF (18 mL) was heated with stirring under N₂ at 85°C for 4 h. After cooling, the insoluble portion was filtered off using a Celite pad and it was washed with DMF (3 x 5 mL). The organic phase was evaporated in vacuo and the residue was chromatographed on silica gel using CH₂Cl₂ - MeOH (98 : 2 → 94 : 6) to give product **16a** (573 mg, 83%), mp 130-132°C. UV max (EtOH) 260 nm (ϵ 14,500), 210 (ϵ 26,100). IR (KBr) 3380 and 3180 (NH₂), 1665, 1620, 1590, 1500, 1265, 1100 cm⁻¹; ¹H NMR (CDCl₃) δ 2.12-2.24 (m, 1 H, H_{4'}), 2.28-2.39 (m, 1 H, H_{2'}), 3.68 (td, 1 H, ²J = ³J = 9.9 Hz, ⁴J_{5'}, F = 1.5 Hz) and 3.84 (ddd, 1 H, ²J = 10.8 Hz, ³J = 6.4 Hz, ⁴J_{5'}, F = 2.0 Hz, H_{5'}), 4.24 (dd, 1 H, ²J = 14.8 Hz, ³J = 8.0 Hz) and 4.54 (ddd, 1 H, partially overlapped with CH₂ of Bn, ²J = 14.8 Hz, ³J = 6.4 Hz, ⁴J_{1'}, F = 3.2 Hz, H_{1'}), 4.53 (AB, 2 H, J_{AB} = 11.6 Hz, CH₂ of Bn), 6.24 (s, 2 H, NH₂), 7.28-7.40 (m, 5 H, C₆H₅), 7.90 and 8.39 (H₂ and H₈ of adenine); ¹³C NMR 25.80 and 26.30 (2t, ²J = 10.4 Hz, C_{2'} and C_{4'}), 38.42 (d, ³J = 5.9 Hz, C_{1'}), 63.35 (d, ³J = 4.5 Hz, C_{5'}), 113.83 (dd, ¹J = 290.3 and 279.9 Hz, C_{3'}); adenine: 120.62 (C₅), 141.20 (C₈), 151.05 (C₄), 154.20 (C₂), 156.81 (C₆); benzyl: 74.21 (CH₂), 128.96 (C_{ortho}), 129.18 (C_{para}), 129.69 (C_{meta}), 138.32 (C_{ipso}); ¹⁹F NMR -124.93 (dt, ²J = 164.9 Hz, ³J_{F, cis-H} = 12.6 Hz), -150.12 (d, ²J = 164.9 Hz). EI-MS 345 (M, 1.6), 325 (M - F - H, 0.8), 296 (M - CF₂ + H, 4.1), 239 (M - OBn + H, 7.7), 224 (M - CH₂OBn, 6.9), 219 (M - OBn - F, 41.8), 148 (AdeCH₂, 10.0), 135 (Ade, 25.5), 91 (Bn, 100.0). CI-MS 346 (M + H, 100.0), 239 (M + H - OBn, 8.6), 219 (M - OBn - F, 13.9), 178 (7.9), 91 (Bn, 10.1). HRMS Calcd. for C₁₇H₁₇F₂N₅O (M): 345.1401. Found: 345.1404. Anal. Calcd. for C₁₇H₁₇F₂N₅O: C, 59.12; H, 4.96; N, 20.28. Found: C, 59.29; H, 5.13; N, 20.45.

(cis)-9-[(2-Hydroxymethyl-3,3-difluorocyclopropyl)methyl]adenine (7a). Boron trichloride (1 M in CH₂Cl₂, 7.4 mL, 7.4 mmol) was added to a solution of compound **16a** (511 mg, 1.48 mmol) in CH₂Cl₂ (23 mL) at -78°C under N₂ over 10 min with stirring. The stirring at -78°C was continued for 5 h whereupon the reaction was quenched by a cautious addition of MeOH (10 mL) and NH₃ in MeOH (10 %, 10 mL). The mixture was allowed to warm up to room temperature, the insoluble portion was filtered off and it was washed with CH₂Cl₂ - MeOH (4 : 1, 4 x 25 mL). The combined organic phases were removed in vacuo and MeOH (2 x 10 mL) was evaporated from the residue. Chromatography on silica gel using CH₂Cl₂ - MeOH (9 : 1 → 4 : 1 containing 0.5 % NH₃) to give product **7a** (354 mg, 93.7 %), mp 198-200°C. UV max (EtOH) 260 nm (ϵ 13,400), 209 (ϵ 20,200). IR (KBr): 3440, 3370 and 3080-3320 (OH, NH₂), 1680, 1620, 1580, 1480, 1310, 1250, 1065, 1030 cm⁻¹; ¹H NMR δ 2.11 (ttd, 1 H, ³J_{cis-H} = ³J_{cis-F} = 12.0 Hz, ³J = 7.6 Hz, ³J_{trans-F} = 1.2 Hz, H_{4'}), 2.47 (tt, 1 H, partially overlapped with d₅-DMSO, ³J_{cis-H} = ³J_{cis-F} = 12.8 Hz, ³J_H = 7.6 Hz, H_{2'}), 3.57-3.65 (m, 1 H) and 3.74 (m, 1 H, J = 12.4 Hz, J = 6.4 Hz, H_{5'}), 4.38 (d, 2 H, ³J = 7.6 Hz, H_{1'}), 5.08 (t, 1 H, ³J = 5.2 Hz, OH), 7.30 (s, 2 H, NH₂), 8.16 and 8.17 (H₂ and H₈ of

adenine). ^{13}C NMR 25.51 (t, $^2J_{\text{F}} = 10.0$ Hz) and 28.70 (t, $^2J_{\text{F}} = 9.3$ Hz, C_2' and C_4'), 38.08 (d, $^3J_{\text{F}} = 5.9$ Hz, C_1'), 55.19 (d, $^3J_{\text{F}} = 5.2$ Hz, C_5'), 115.80 (dd, $^1J_{\text{F}} = 288.5$ Hz and 281.1 Hz, C_3'); adenine: 120.15 (C_5), 142.04 (C_8), 150.86 (C_4), 154.08 (C_2), 157.54 (C_6). ^{19}F NMR -118.90 (dt, $^2J_{\text{F}} = 160.3$ Hz, $^3J_{\text{cis-H}} = 12.8$ Hz), -145.55 (d, $^2J_{\text{F}} = 160.3$ Hz). EI-MS 255 (M , 21.0), 238 ($\text{M} + \text{H} - \text{H}_2\text{O}$, 12.0), 235 ($\text{M} - \text{H} - \text{F}$, 15.4), 224 ($\text{M} - \text{CH}_2\text{OH}$, 8.4), 206 ($\text{M} + \text{H} - \text{CH}_2\text{OH} - \text{F}$, 41.5), 149 (AdeCH_2 , 24.0), 135 (adenine, 100.0), 108 ($\text{M} + \text{H} - \text{AdeCH}_2$, 37.1). HRMS Calcd. for $\text{C}_{10}\text{H}_{11}\text{F}_2\text{N}_5\text{O}$ (M): 255.0932. Found: 255.0931. Anal. Calcd. for $\text{C}_{10}\text{H}_{11}\text{F}_2\text{N}_5\text{O}$: C, 47.06; H, 4.34; N, 27.44; F, 14.90. Found: C, 47.25; H, 4.55; N, 27.33; F, 14.71.

(*cis*)-2-Amino-9-[(2-benzyloxymethyl-3,3-difluorocyclopropyl)methyl]-6-chloropurine (16b).

The reaction and workup followed the procedure described for adenine analogue **16a**. A mixture of 2-amino-6-chloropurine (678 mg, 4 mmol), compound **8** (873 mg, 3 mmol) and K_2CO_3 (1.10 g, 8 mmol) in DMF (22 mL) was stirred under N_2 at 85°C for 5 h to give after chromatography on silica gel using CH_2Cl_2 - MeOH (99 : 1 \rightarrow 97 : 3) product **16b** (788 mg, 61.6 %), mp $130\text{--}133^\circ\text{C}$. UV max (EtOH) 310 nm (ϵ 7,700), 248 (ϵ 6,500), 222 (ϵ 28,100). IR (KBr) 3460, 3360, and 3240 (NH_2), 1650, 1620, 1580, 1480, 1420, 1220, 1080 cm^{-1} . ^1H NMR (CDCl_3) δ 2.12–2.24 (m, 1 H, $J = 12.8$ Hz, $J = 6.8$ Hz, H_4'), 2.24–2.35 (m, 1 H, $J = 8.0$ Hz, $J = 7.2$ Hz, H_2'), 3.64 (t, 1 H, $^2J = ^3J = 10.0$ Hz) and 3.84 (ddd, 1 H, $^2J = 10.8$ Hz, $^3J = 6.4$ Hz, $^4J_{\text{F}} = 2.0$ Hz, H_5'), 4.13 (dd, 1 H, $^2J = 14.8$ Hz, $^3J = 8.0$ Hz) and 4.38 (ddd, 1 H, $^2J = 14.8$ Hz, $^3J = 6.8$ Hz, $^4J_{\text{F}} = 3.2$ Hz, H_1'), 4.53 (AB, 2 H, $J_{\text{AB}} = 11.6$ Hz, CH_2 of Bn), 5.40 (s, 2 H, NH_2), 7.29–7.40 (m, 5 H, Ph), 7.85 (Hg of purine). ^{13}C NMR 25.47 (t, $^2J_{\text{F}} = 10.4$ Hz) and 26.28 (t, $^2J_{\text{F}} = 10.4$ Hz, C_2' and C_4'), 38.42 (d, $^3J_{\text{F}} = 6.7$ Hz, C_1'), 63.28 (d, $^3J_{\text{F}} = 4.4$ Hz, C_5'), 113.72 (dd, $^1J_{\text{F}} = 290.4$ Hz, $^1J_{\text{F}} = 280.0$ Hz, C_3'); purine: 126.22 (C_5), 143.07 (C_8), 152.46 (C_4), 154.78 (C_2), 160.27 (C_6); benzyl: 74.27 (CH_2), 128.99 (C_{ortho}), 129.24 (C_{para}), 129.72 (C_{meta}), 138.20 (C_{ipso}). ^{19}F NMR (CDCl_3 , 376 MHz): -124.82 (dt, $^2J_{\text{F}} = 164.9$ Hz, $^3J_{\text{cis-H}} = 12.6$ Hz), -149.90 (d, $^2J_{\text{F}} = 164.9$ Hz). EI-MS m/z : 379 and 381 (M , 7.1, 2.6), 288 and 290 ($\text{M} - \text{Bn}$, 1.3, 0.5), 273 and 275 ($\text{M} - \text{OBn} + \text{H}$, 10.8, 3.4), 253 and 255 ($\text{M} - \text{OBn} - \text{F}$, 54.4, 18.0), 240 and 242 ($\text{M} - \text{CH}_2\text{OBn} - \text{F} + \text{H}$, 5.3, 1.6), 183 and 185 (2-amino-6-chloropurine- $\text{CH}_2 + \text{H}$, 6.5, 2.2), 169 and 171 (2-amino-6-chloropurine, 22.7, 7.6), 91 (Bn, 100.0). HRMS Calcd. for $\text{C}_{17}\text{H}_{16}^{35}\text{ClF}_2\text{N}_5\text{O}$ (M): 379.1011. Found: 379.1006.

(*cis*)-2-Amino-9-[(2-hydroxymethyl-3,3-difluorocyclopropyl)methyl]-6-chloropurine (7e).

Boron trichloride (1.0 M in CH_2Cl_2 , 20.6 mL, 20.6 mmol) was added to a solution of compound **16b** (782 mg, 2.06 mmol) in CH_2Cl_2 (65 mL) at -78°C under N_2 during 10 min with stirring. The stirring at -78°C was continued for 7 h whereupon the reaction was quenched by addition of MeOH (10 mL) and solid NaHCO_3

(10 g, 119 mmol) with stirring. The mixture was cautiously warmed up to room temperature (**caution**: foaming between -30 and -20°C!). The solids were filtered off using a Celite pad and they were washed with CH₂Cl₂ - MeOH (95 : 5, 3 x 50 mL). The solvents were removed in vacuo and MeOH (2 x 10 mL) was evaporated from the residue. The crude product was chromatographed on silica gel using CH₂Cl₂ - MeOH (97 : 3 → 92 : 8) to give compound **7e** (537 mg, 90 %), mp 165-167°C with resolidification and melting at 180-182°C. UV max (EtOH) 310 nm (ϵ 8,100), 248 (ϵ 6,600), 223 (ϵ 29,600). IR (KBr) 3530, 3350 and 3220 (OH, NH₂), 1630, 1580, 1480, 1420, 1295, 1220, 1010, 935, 920 cm⁻¹. ¹H NMR δ 2.20 (ttd, 1 H, ³J_{cis-H} = ³J_{cis-F} = 12.0 Hz, ³J_H = 7.8 Hz, ³J_{trans-F} = 1.6 Hz, H_{4'}), 2.47 (tt, 1 H, partially overlapped with d₅-DMSO, ³J_{cis-H} = ³J_{cis-F} = 12.8 Hz, ³J_H = 8.0 Hz, H_{2'}), 3.64-3.70 (m, 1 H) and 3.82 (m, 1 H, apparent J = 12.4 Hz, J = 5.2 Hz, H_{5'}), 4.38 (d, 2 H, ³J = 8.0 Hz, H_{1'}), 5.06 (t, 1 H, ³J = 5.2 Hz, OH), 7.07 (s, 2 H, NH₂), 8.23 (Hg of purine). ¹³C NMR 25.16 (t, ²J_F = 10.1 Hz) and 28.76 (t, ²J_F = 9.3 Hz, C_{2'} and C_{4'}), 38.23 (d, ³J_F = 6.0 Hz, C_{1'}), 55.24 (d, ³J_F = 5.2 Hz, C_{5'}), 115.85 (dd, ¹J_F = 289.3 Hz and 280.4 Hz, C_{3'}); purine: 124.84 (C₅), 144.35 (C₈), 151.04 (C₄), 155.58 (C₂), 161.46 (C₆). ¹⁹F NMR -123.76 (dt, ²J_F = 160.3 Hz, ³J_{cis-H} = 13.0 Hz), -150.24 (d, ²J_F = 160.3 Hz). EI-MS 289 and 291 (M, 44.5, 14.8), 272 and 274 (M + H - H₂O, 7.9, 2.6), 240 and 242 (M + H - CH₂OH - F, 10.6, 3.3), 182 and 184 (2-amino-6-chloropurine-CH₂, 10.1, 4.9), 169 and 171 (2-amino-6-chloropurine, 100.0, 33.1). HRMS Calcd. for C₁₀H₁₀ClF₂N₅O (M): 289.0542. Found: 289.0541. Anal. Calcd. for C₁₀H₁₀³⁵ClF₂N₅O: C, 41.46; H, 3.48; N, 24.18; Cl, 12.24. Found: C, 41.51; H, 3.60; N, 24.33; Cl, 12.34.

(*cis*)-**9-[(2-Hydroxymethyl-3,3-difluorocyclopropyl)methyl]guanine (7b)**. A solution of compound **7e** (232 mg, 0.8 mmol) in formic acid (80 %, 10 mL) was heated at 80°C for 3 h with stirring. Volatile components were removed in vacuo leaving a sirup which was dissolved in NH₃/MeOH (20 %, 10 mL). The mixture was stirred at room temperature for 18 h. After evaporation, MeOH was added and the product **7b** was filtered off (207 mg, 95 %), mp 296-300°C (decomp.). UV max (EtOH) 254 nm (ϵ 15,200), 204 (ϵ 17,800). IR (KBr) 3520, 3320 and 3130 (OH, NH₂), 1755, 1700, 1650, 1600, 1560, 1490, 1305, 1195, 1050, 1010 cm⁻¹. ¹H NMR δ 2.06 (tt, 1 H, ³J_{cis-H} = ³J_{cis-F} = 12.2 Hz, ³J_H = 8.0 Hz, H_{4'}), 2.36 (tt, 1 H, ³J_{cis-H} = ³J_{cis-F} = 12.2 Hz, ³J_H = 8.0 Hz, H_{2'}), 3.53 (poorly resolved dd, 1 H, ²J = 9.2 Hz, ³J = 8.8 Hz) and 3.82 (m, 1 H, ²J = 12.4 Hz, ³J = 5.2 Hz, H_{5'}), 4.13 (AB, 2 H, J_{AB} = 16.4 Hz, H_{1'}), 4.94 (br s, 1 H, OH), 6.47 (s, 2 H, NH₂), 7.67 (Hg), 10.59 (CONH). ¹³C NMR 24.39 (t, ²J_F = 10.4 Hz) and 27.56 (t, ²J_F = 9.3 Hz, C_{2'} and C_{4'}), 36.73 (d, ³J_F = 6.4 Hz, C_{1'}), 54.14 (d, ³J_F = 5.2 Hz, C_{5'}), 114.81 (dd, ¹J_F = 287.7 Hz and 280.4 Hz, C_{3'}); purine: 116.93 (C₅), 137.40 (C₈), 151.57 (C₄), 154.14 (C₂), 157.30 (C₆). ¹⁹F NMR -123.84 (dt, ²J_F = 160.3 Hz, ³J_{cis-H} = 13.4 Hz), -150.50 (d, ²J_F = 160.3 Hz). FAB-MS (TG) 543 (2M + H, 4.7),

272 (M + H, 100.0), 181 (7.8), 152 (guanine + H, 27.3). Anal. Calcd for C₁₀H₁₁F₂N₅O₂: C, 44.28; H, 4.09; N, 25.82; F, 14.01. Found: C, 44.75; H, 4.33; N, 26.12; F, 13.96.

(*cis*)-1-[(2-Benzoyloxymethyl-3,3-difluorocyclopropyl)methyl]cytosine

(**17**). A mixture of N⁴-acetylcytosine (612 mg, 4 mmol), compound **8** (728 mg, 2.5 mmol) and K₂CO₃ (0.97 g, 7.0 mmol) in DMF (20 mL) was stirred under N₂ at 85°C for 3 h. Water (7.5 mL) was then added and temperature was kept at 85°C for another 2 h. After cooling, the solids were filtered off through a Celite pad and they were washed with DMF (5 x 6 mL). The combined filtrate and washings were evaporated. The residue was treated with a mixture of CH₂Cl₂ - MeOH (4 : 1, 50 mL), the insoluble portion was filtered off and it washed with the same solvent (2 x 30 mL). The organic phase was evaporated and the residue was chromatographed on silica gel using CH₂Cl₂ - MeOH (95 : 5 → 85 : 15) to give product **17** (564 mg, 70 %), mp 215-218°C. UV max (EtOH) 273 nm (ε 8,700), 242 (shoulder, ε 7,300), 205 (ε 26,200). IR (KBr) 3370 and 3140 (NH₂), 1665, 1630, 1485, 1390, 1285, 1080 cm⁻¹. ¹H NMR δ 2.23 (tt, 1 H, ³J_{cis-H} = ³J_{cis-F} = 12.8 Hz, ³J_H = 6.8 Hz, H_{4'}), 2.32 (tt, 1 H, ³J_{cis-H} = ³J_{cis-F} = 14.0 Hz, ³J_H = 6.8 Hz, H_{2'}), 3.63-3.73 (m, 2 H, J = 9.2 Hz, J = 7.6 Hz, H_{5'}), 3.73 (dd, 1 H, ²J = 14.4 Hz, ³J = 7.6 Hz) and 3.97 (ddd, 1 H, ²J = 14.4 Hz, ³J = 6.8 Hz, ⁴J_F = 2.8 Hz, H_{1'}), 4.50 (s, 2 H, CH₂ of Bn), 5.65 (d, 1 H, ³J = 7.2 Hz, H₅), 7.13 (d, 2 H, ²J = 24.8 Hz, NH₂), 7.53 (d, 1 H, ³J = 7.2 Hz, H₆), 7.28-7.40 (m, 5 H, C₆H₅). ¹³C NMR 25.28 (t, ²J_F = 9.6 Hz) and 26.00 (t, ²J_F = 8.9 Hz, C_{2'} and C_{4'}), 43.93 (d, ³J_F = 5.5 Hz, C_{1'}), 64.02 (d, ³J_F = 4.7 Hz, C_{5'}), 116.02 (dd, ¹J_F = 288.4 Hz and 280.4 Hz, C_{3'}); cytosine: 95.20 (C₅), 147.13 (C₆), 157.29 (C₂), 167.59 (C₄); benzyl: 73.36 (CH₂), 129.11 (C_{para}), 129.19 (C_{ortho}), 129.85 (C_{meta}), 139.58 (C_{ipso}). ¹⁹F NMR -118.63 (dt, ²J_F = 158.5 Hz, ³J_{cis-H} = 13.0 Hz), -143.99 (d, ²J_F = 158.8 Hz). EI-MS 321 (M, 2.5), 230 (M - Bn, 1.9), 215 (M + H - OBn, 38.7), 195 (M - OBn - F, 38.7), 136 (19.5), 111 (cytosine, 67.5), 91 (Bn, 100.0). HRMS Calcd. for C₁₆H₁₇F₂N₃O₂ (M): 321.1289. Found: 321.1292. Anal. Calcd. for C₁₆H₁₇F₂N₃O₂: C, 59.81; H, 5.33; N, 13.08. Found: C, 59.88; H, 5.18; N, 13.22.

(*cis*)-1-[(2-Hydroxymethyl-3,3-difluorocyclopropyl)methyl]cytosine (7c).

Benzoic anhydride was added to a stirred and refluxing solution of compound **17** (400 mg, 1.24 mmol) in EtOH (40 mL) in six increments of 282 mg (1.25 mmol) each hour (total 1.69 g, 7.5 mmol). The mixture was then refluxed for another hour. After cooling, it was evaporated and the residue was partitioned between CH₂Cl₂ (80 mL) and Na₂CO₃ (5 %, 80 mL). The aqueous phase was extracted with CH₂Cl₂ (3 x 30 mL). The organic phase was washed with saturated NaCl (50 mL)/NaHCO₃ (20 mL) and it was dried (Na₂SO₄). After evaporation, the residue was chromatographed on silica gel using CH₂Cl₂ - MeOH

(95 : 5) to give the N⁴-benzoyl derivative **18** (520 mg, 98%), mp 160-162°C. UV max (EtOH) 305 nm (ϵ 9,800), 259 (ϵ 23,600), 205 (ϵ 31,700).

Compound **18** (515 mg, 1.21 mmol) was debenzylated as described for 2-amino-6-chloropurine derivative **16b**. A solution of the crude product **7c** in NH₃/MeOH (20 %, 30 mL) was stirred at room temperature for 18 h. Volatile components were evaporated and the residue was chromatographed on silica gel using CH₂Cl₂ - MeOH (4 : 1 containing 0.5 % NH₃) to give compound **7c** (266 mg, 95 %), mp 237-240°C. UV max (EtOH) 273 nm (ϵ 8,100), 203 (ϵ 20,100). IR (KBr) 3400 and 3140 (OH and NH₂), 1665, 1620, 1490, 1400, 1040 cm⁻¹. ¹H NMR δ 2.00 (tt, 1 H, ³J_{cis-H} = ³J_{cis-F} = 12.4 Hz, ³J_H = 7.6 Hz, H_{4'}), 2.16 (tt, 1 H, ³J_{cis-H} = ³J_{cis-F} = 12.4 Hz, ³J_H = 7.2 Hz, H_{2'}), 3.51-3.65 (m, 2 H, H_{5'}), 3.68 (dd, 1 H, ²J = 14.4 Hz, ³J = 7.6 Hz) and 3.95 (ddd, 1 H, ²J = 14.0 Hz, ³J = 6.8 Hz, ⁴J_F = 3.2 Hz, H_{1'}), 4.95 (br, 1 H, OH), 5.67 (d, 1 H, ³J = 7.2 Hz, H₅), 7.14 (d, 2 H, ²J = 28.8 Hz, NH₂), 7.53 (d, 1 H, ³J = 7.2 Hz, H₆). ¹³C NMR 24.10 (t, ²J_F = 10.0 Hz) and 27.65 (t, ²J_F = 10.4 Hz, C_{2'} and C_{4'}), 42.91 (d, ³J_F = 4.2 Hz, C_{1'}), 54.26 (d, ³J_F = 5.2 Hz, C_{5'}), 115.19 (dd, ¹J_F = 287.7 Hz and 280.4 Hz, C_{3'}); cytosine: 94.21 (C₅), 146.23 (C₆), 156.17 (C₂), 166.43 (C₄). ¹⁹F NMR -123.29 (dt, ²J_F = 158.8 Hz, ³J_{cis-H} = 13.7 Hz), -149.82 (d, ²J_F = 158.8 Hz). EI-MS 231 (M, 7.3), 214 (M + H - H₂O, 100.0), 200 (M - CH₂OH, 83.8), 182 (M + H - CH₂OH - F, 15.7). HRMS Calcd. for C₉H₁₁F₂N₃O₂ (M): 231.0819. Found: 231.0822. Anal. Calcd. for C₉H₁₁F₂N₃O₂: C, 46.76; H, 4.80; N, 18.17; F, 16.43. Found: C, 46.50; H, 5.04; N, 17.97; F, 16.23.

(*cis*)-1-[(2-Benzylloxymethyl-3,3-difluorocyclopropyl)methyl]thymine (**19**) and 1-Benzylthymine (**20**). A solution of compound **8** (728 mg, 2.5 mmol) and 2,4-bis-O-(trimethylsilyl)-5-methylpyrimidine (1.35 g, 5.0 mmol) in acetonitrile (5 mL) was refluxed with stirring under N₂ for 7 days. After cooling, methanol (2 mL) was added, the precipitate was removed by centrifugation and it was washed with CH₂Cl₂ (3 x 30 mL). The organic phase was evaporated and the residue was chromatographed on silica gel using CH₂Cl₂ - MeOH (98 : 2 \rightarrow 96 : 4) to give an inseparable mixture of product **19** and 1-benzylthymine (**20**) in the ratio of 1.5 : 1 based on ¹H NMR analysis (714 mg, 51 %) as a foam. UV max (EtOH) 270 and 208 nm. IR (KBr) 3200 and 3080 (NH), 1690, 1480, 1380, 1260, 1090 cm⁻¹; ¹H NMR (CDCl₃), compound **19**, δ 1.92 (d, 3 H, ⁴J = 1.2 Hz, 5-CH₃), 2.19-2.28 (m, 2 H, J = 8.0 Hz, J = 4.4 Hz, H_{2'} and H_{4'}), 3.68-3.75 (m, 2 H, H_{5'}), 3.86-3.94 (m, 1 H, J = 4.4 Hz, J = 2.0 Hz) and 4.26 (dm, 1 H, ²J = 14.4 Hz, J = 5.6 Hz, J = 3.6 Hz, H_{1'}), 4.64 (AB, 2 H, J_{AB} = 12.0 Hz, CH₂ of Bn), 7.16 (q, 1 H, ⁴J = 1.2 Hz, H₆), 7.38-7.50 (m, 5 H, C₆H₅), 9.83 (s, 1 H, NH); 1-benzylthymine (**20**), 1.99 (d, 3 H, ⁴J = 1.2 Hz, 5-CH₃), 5.01 (s, CH₂ of Bn), 7.10 (q, 1 H, ⁴J = 1.2 Hz, H₆), 7.38-7.50 (m, 5 H, Ph), 9.83 (s, 1 H, NH). ¹³C NMR, compound **19**, 25.29 (t, ²J_F = 10.4 Hz) and 26.29 (t, ²J_F = 10.0 Hz, C_{2'} and C_{4'}), 43.18 (d, ³J_F = 5.2 Hz,

C1'), 63.52 (d, $^3J_F = 4.4$ Hz, C5'), 114.15 (dd, $^1J_F = 290.8$ Hz and 279.6 Hz, C3'); benzyl: 74.28 (CH₂), 129.06 (Cortho), 129.29 (Cpara), 129.80 (Cmeta), 138.43 (Cipso); thymine: 13.49 (5-CH₃), 112.47 (C5), 141.00 (C6), 152.29 (C2), 165.60 (C4); 1-benzylthymine (**20**), benzyl: 52.12 (CH₂), 129.19 (Cortho), 129.62 (Cpara), 130.28 (Cmeta), 136.73 (Cipso); thymine: 13.63 (5-CH₃), 112.49 (C5), 141.00 (C6), 152.59 (C2), 165.65 (C4). ^{19}F NMR, compound **19**, -129.42 (dt, $^2J_F = 166.4$ Hz, $^3J_{cis-H} = 11.5$ Hz), -153.73 (d, $^2J_F = 164.5$ Hz).

(cis)-1-((2-Hydroxymethyl-3,3-difluoro)cyclopropylmethyl)thymine (7d).

The procedure followed that described for the adenine analog **7a**. Thus, a mixture of compound **19** and 1-benzylthymine (**20**) from the previous experiment in CH₂Cl₂ (10 mL) was treated with BCl₃ (1.0 M in CH₂Cl₂, 24.3 mmol) at -78°C for 7 h to give 1-benzylthymine (**20**, 190 mg, 90.5%) and product **7d** (335 mg, 93 %) after chromatography on silica gel using CH₂Cl₂ - MeOH (97 : 3 → 9 : 1). 1-Benzylthymine (**20**), mp 154-155°C after recrystallization from EtOH; lit.²¹ 161-163°C. UV max (EtOH) 271 nm (ϵ 10,700), 207 (ϵ 17,800); lit.²¹ UV max (MeOH) 271 nm (ϵ 10,500). For 1H and ^{13}C NMR see the previous experiment; the 1H NMR chemical shifts were similar to those reported¹⁹ for oily product **20**. EI-MS 216 (M, 12.9), 91 (Bn, 100.0); HRMS Calcd. for C₁₂H₁₂N₂O₂ (M): 216.0899. Found: 216.0901.

Compound **7d**, mp 178-180°C. UV max (EtOH) 268 nm (ϵ 10,000), 206 (ϵ 9,800). IR (KBr) 3460, 3200 and 3060 (OH, NH), 1710, 1650, 1490, 1360, 1260, 1070, 1010 cm⁻¹; 1H NMR 1.73 (d, 3 H, $^4J = 1.2$ Hz, 5-CH₃), 2.02 (ttd, 1 H, $^3J_{cis-H} = ^3J_{cis-F} = 12.8$ Hz, $^3J_H = 8.0$ Hz, $^3J_{trans-F} = 1.4$ Hz, H4'), 2.18 (tt, 1 H, $^3J_{cis-H} = ^3J_{cis-F} = 12.4$ Hz, $^3J_H = 7.6$ Hz, H2'), 3.49-3.58 (m, 1 H) and 3.63 (m, 1 H, H5'), 3.73 (dd, 1 H, $^2J = 14.4$ Hz, $^3J = 8.0$ Hz) and 3.92 (ddd, 1 H, $^2J = 14.4$ Hz, $^3J = 7.2$ Hz, $^4J_F = 3.2$ Hz, H1'), 4.91 (t, 1 H, $^3J = 5.4$ Hz, OH), 7.47 (d, 1 H, $^4J = 1.2$ Hz, H6), 11.29 (NH). ^{13}C NMR 23.81 (t, $^2J_F = 10.0$ Hz) and 27.62 (t, $^2J_F = 9.3$ Hz, C2' and C4'), 41.34 (d, $^3J_F = 5.2$ Hz, C1'), 54.26 (d, $^3J_F = 5.9$ Hz, C5'), 114.95 (dd, $^1J_F = 288.5$ Hz and 280.3 Hz, C3'); thymine: 12.54 (5-CH₃), 109.32 (C5), 141.35 (C6), 151.33 (C2), 164.71 (C4); ^{19}F NMR -123.40 (dt, $^2J_F = 158.8$ Hz, $^3J_{cis-H} = 13.0$ Hz), -149.50 (d, $^2J_F = 158.8$ Hz). EI-MS 246 (M, 30.2), 229 (M + H - H₂O, 69.2), 215 (M - CH₂OH, 61.8), 197 (M + H - CH₂OH - F, 10.3), 172 (23.9), 149 (41.4), 139 (thymine-CH₂, 35.1), 126 (thymine, 66.4), 96 (100.0). HRMS Calcd. for C₁₀H₁₂F₂N₂O₃ (M): 246.0816. Found: 246.0815. Anal. Calcd. for C₁₀H₁₂F₂N₂O₃: C, 48.78; H, 4.91; N, 11.38; F, 15.43. Found: C, 49.00; H, 5.10; N, 11.47; F, 15.27.

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REFERENCES

1. Welch, J. T.; Eswarakrishnan, S. *Fluorine in Bioorganic Chemistry*, John Wiley & Sons, New York, **1991**, pp. 220-233.
2. Herdewijn, P.; Van Aerschot, A.; Kerremans, L. *Nucleosides & Nucleotides* **1989** *8*, 65-96.
3. Huryn, D. M.; Okabe, M. *Chem. Rev.* **1992** *92*, 1745-1768.
4. Hertel, L. W.; Boder, G. B.; Kroin, J. S.; Rinzel, S. M.; Poore, G. A.; Todd, G. C.; Grindey, G. B. *Cancer Res.* **1990** *50*, 4417-4422.
5. Halazy, S.; Ehrhard, A.; Danzin, C. *J. Am. Chem. Soc.* **1991** *113*, 315-317.
6. Casara, P. J.; Kenny, M. T.; Jund, K. C. *Tetrahedron Lett.* **1991** *32*, 3823-3826.
7. Sekiyama, T.; Hatsuya, S.; Tanaka, Y.; Uchiyama, M.; Ono, N.; Iwayama, S.; Oikawa, M.; Suzuki, K.; Okunishi, M.; Tsuji, T. *J. Med. Chem.* **1998** *41*, 1284-1298.
8. Ashton, W. T.; Meurer, L. C.; Cantone, C. L.; Field, A. K.; Hannah, J.; Karkas, J. D.; Liou, R.; Patel, G. F.; Perry, H. C.; Wagner, A. F.; Walton, E.; Tolman, R. L. *J. Med. Chem.* **1988** *31*, 2304-2315.
9. Czernecki, S.; Georgoulis, C.; Provelenghiou, C. *Tetrahedron Lett.* **1976**, 3535-3536.
10. Danishefsky, S.; Regan, J. *Tetrahedron Lett.* **1981** *22*, 3919-3922.
11. Naruta, Y.; Nagai, N.; Maruyama, K. *Chem. Lett.* **1983**, 1383-1386.
12. Corey, E. J.; Venkateswarlu, A. *J. Am. Chem. Soc.* **1972** *94*, 6190-6191.
13. Taguchi, T.; Kurishita, M.; Shibuya, A.; Aso, K. *Tetrahedron* **1997** *53*, 9497-9508.
14. Verheyden, J. P. H.; Moffatt, J. G. *J. Org. Chem.* **1972** *37*, 2289-2299.
15. Qiu, Y.-L.; Ksebati, M.; Ptak, R. G.; Fan, B. Y.; Breitenbach, J. M.; Lin, J.-S.; Cheng, Y.-C.; Kern, E. R.; Drach, J. C.; Zemlicka, J. *J. Med. Chem.* **1998** *41*, 10-23.
16. Qiu, Y.-C.; Ptak, R. G.; Breitenbach, J. M.; Lin, J.-S.; Cheng, Y.-C.; Kern, E. R.; Drach, J. C.; Zemlicka, J. *Antiviral Chem. Chemother.* **1998** *9*, 341-352.
17. Xu, Z.-Q.; Joshi, R. V.; Zemlicka, J. *Tetrahedron* **1995** *51*, 67-76.

18. Jones, B. C. N. M.; Silverton, J. V.; Simons, C.; Megati, S.; Nishimura, H.; Maeda, Y.; Mitsuya, H.; Zemlicka, J. *J. Med. Chem.* **1995** *38*, 1397-1405.
19. Koomen, G. J.; Provoost, L. M.; van Maarschalkerwaart, D. A. H.; Willard, N. P. *Nucleosides & Nucleotides* **1992** *11*, 1297-1303.
20. Phadtare, S.; Kessel, D.; Corbett, T. H.; Renis, H. E.; Court, B. A.; Zemlicka, J. *J. Med. Chem.* **1991** *34*, 421-429.
21. Smith, R. C.; Binkley, S. B. *J. Org. Chem.* **1959** *24*, 249-251.

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